
AMERICAN JOURNAL OF DENTISTRY

Volume 28, No. 6, December, 2015 - p. 309-376

EDITOR

Franklin García-Godoy

MANAGING EDITOR

Katherine J. García-Godoy

EDITORIAL BOARD

Michael C. Alfano

Thomas Attin

Stephen Bayne

Daniel C.N. Chan

Gordon J. Christensen

Sebastian G. Ciancio

Gary A. Crim

Jaime Cury

Kevin J. Donly

Frederick Eichmiller

Albert J. Feilzer

Jack L. Ferracane

Marco Ferrari

Catherine M. Flaitz

Roland Frankenberger

Robert W. Gerlach

Reinhard Hickel

M. John Hicks

Mark E. Jensen

Andrej M. Kielbassa

Norbert Krämer

Ivo Krejci

Grayson W. Marshall

Sally J. Marshall

John F. McCabe

Peter E. Murray

Raquel Osorio

Cornelis H. Pameijer

Rade D. Paravina

Jorge Perdigão

John M. Powers

Mark S. Putt

Sol Silverman

Karl-Johan Söderholm

Hans-Jörg Staehle

Edward J. Swift, Jr.

Junji Tagami

Franklin Tay

Manuel Toledano

Bart Van Meerbeek

Anthony R. Volpe

Ann Wennerberg

Donald J. White

STATISTICAL CONSULTANT

Daniel L. Jones

AMERICAN JOURNAL OF DENTISTRY

Published By Mosher & Linder, Inc.

Volume 28, Number 6, December, 2015 - p. 309 - 376

www.amjdent.com

CONTENTS

Review Articles

Surface properties of resin-based composite materials and biofilm formation:
A review of the current literature.
G. Cazzaniga, M. Ottobelli, A. Ionescu, F. Garcia-Godoy & E. Brambilla

311

Adhesive sealing of dentin surfaces in vitro: A review.
M.M. Abu Nawareg, A.Z. Zidan, J. Zhou, A. Chiba, J. Tagami & D.H. Pashley

321

Research Articles

Dentin wear after simulated toothbrushing with water, a liquid dentifrice or a standard toothpaste.
Y. Jang, J-J. Ihm, S-J. Baik, K-J. Yoo, D-H. Jang, B-D. Roh & D-G. Seo

333

Evaluation of two disinfection/sterilization methods on silicon rubber-based composite finishing instruments.
V.A. Lacerda, L.O. Pereira, R. Hirata Junior & C.R. Perez

337

Efficacy of different “in-office” desensitizing treatment methods:
An in vitro SEM analysis.
R. De Cássia Gomes Camacho, G. Lecio Miranda, F. Tonasso Oliveira,
F. Vieira Ribeiro, S. Peres Pimentel & R. Corrêa Viana Casarin

342

Effect of sonic vibration of an ultrasonic toothbrush on the removal of *Streptococcus mutans* biofilm from enamel surface.
L.N. Hashizume & A. Dariva

347

Randomized controlled trial comparing a powered toothbrush with distinct multi-directional cleaning action to a manual flat trim toothbrush.
J. Gallob, L.R. Mateo, P. Chaknis, B.M. Morrison Jr & F. Panagakos

351

Depth of cure of bulk fill composites with monowave and polywave curing lights.
T.S. Menees, C.P. Lin, D.D. Kojic, J.O. Burgess & N.C. Lawson

357

Two pre-treatments for bonding to non-carious cervical root dentin.
S. Flury, A. Peutzfeldt & A. Lussi

362

Influence of organic acids present in oral biofilm on the durability of the repair bond strength, sorption and solubility of resin composites.
S. da Silva, E. Moreira da Silva, M.B. Ferreira Delphim, L.T. Poskus & C. Mariote Amaral

367

2015 Index of the American Journal of Dentistry

373

Dentin wear after simulated toothbrushing with water, a liquid dentifrice or a standard toothpaste

YOUNGJUNE JANG, DDS, JUNG-JOON IHM, PHD, SU-JIN BAIK, KYUNG-JIN YOO, DA-HYUN JANG, BYOUNG-DUCK ROH, DDS, MSD, PHD & DEOG-GYU SEO, DDS, MSD, PHD

ABSTRACT: Purpose: To investigate the influence of dentifrices with and without abrasives on the wear and surface topography of human dentin following simulated toothbrushing in vitro. **Methods:** 24 dentin specimens were prepared and randomly allocated to a liquid dentifrice (Garglin Gum-Guard), conventional dentifrice (333 Clinic Total Care), and control (distilled water) groups. Specimens were subjected to simulated toothbrushing of 50,000 repeated strokes under a 150 g-load. The dentin surface was profiled in each specimen using a profilometer before and after toothbrushing. The mean surface roughness (Ra) of the specimens was calculated and compared by one-way ANOVA and Tukey's post-hoc test ($\alpha=0.05$). The dentin surfaces were further examined by scanning electron microscopy (SEM). **Results:** The Ra values were similar between the liquid dentifrice and control groups ($P>0.05$), and was significantly higher in the conventional dentifrice group ($P<0.001$). On SEM examination, patent dentin tubules were observed in the conventional dentifrice and liquid dentifrice groups, but were not observed in the control group. (*Am J Dent* 2015;28:333-336).

CLINICAL SIGNIFICANCE: Liquid dentifrice reduced dentin wear caused by toothbrushing compared to conventional dentifrice. However, for dentin hypersensitivity, liquid dentifrice requires further validation because both conventional and liquid dentifrices caused patent dentin tubules on the worn dentin surface.

✉: Dr. Deog-Gyu Seo, Department of Conservative Dentistry and Dental Research Institute, School of Dentistry, Seoul National University, 101 Daehak-ro, Jongno-gu, Seoul, Korea. E-mail: dgseo@snu.ac.kr

Introduction

Dentin hypersensitivity is one of the most common dental issues, comprising more than one-quarter of all cases of oral and maxillofacial pain in adults.^{1,2} Although the mechanism of dentin hypersensitivity is not fully understood, the non-caries cervical lesion (NCCL), including cervical abrasion, is a major contributing factor.^{3,4} Daily toothbrushing is important in the development of dentin hypersensitivity because toothbrushing is reportedly closely associated with NCCL.^{5,6} Toothbrushing alone rarely causes excessive wear to the dental hard tissue,^{6,9} but when it is combined with the abrasives in dentifrice, cervical dentin wear may be accelerated.¹⁰

Conventional dentifrices contain a variety of abrasives such as silica, alumina, dicalcium phosphate dihydrate, and calcium carbonate to improve their cleaning power, which may increase dentin wear.¹¹ In this context, a recently introduced liquid dentifrice, which does not contain dental abrasives, may have clinical potential to reduce dentin wear caused by daily toothbrushing. Turssi et al⁹ reported that the magnitude of dentin wear from toothbrushing was associated with the concentration of abrasives in the dentifrice; thus, liquid dentifrice appears to be advantageous compared with conventional dentifrices in reducing NCCL development. However, evidence concerning the wear characteristics of liquid dentifrice is still lacking.

The present study investigated the effect of liquid dentifrice on the wear and surface topography of human dentin under simulated toothbrushing in vitro. The null hypothesis in this study was the following: liquid dentifrice and conventional dentifrice did not differ in the magnitude of wear and surface morphology alteration in human dentin when used in conjunction with toothbrushing.

Materials and Methods

Dentin specimen preparation - This study was approved by the Research Ethics Committee of Seoul National University,

Graduate School of Dentistry (IRB number: S-D20140001). The specimen preparation, dentin wear test, and measurements were performed in accordance with the International Organization for Standardization (ISO) 11609 standards.

A commercial conventional dentifrice (333 Clinic Total Care^a) and a liquid dentifrice (Garglin Gum-Guard^a) were used for the two experimental groups, and sterile distilled water^b served as the negative control. The composition of each product is described in Table 1.

Twelve sound maxillary and mandibular premolars without dental caries or abrasion were extracted and stored in 0.1% thymol solution until specimen preparation. A total 4 mm of cervical dentin centered at the cemento-enamel junction (CEJ) was isolated from each tooth (2 mm each on the occlusal and apical sides of the CEJ). Cylindrical dentin blocks were generated by removing the occlusal and radicular tooth portions with a diamond disk. The dentin blocks were sectioned sagittally to produce two symmetrical dentin specimens from each tooth (Fig. 1).

Of the 24 dentin specimens, the 16 specimens obtained from the eight premolars were randomly allocated into the two experimental groups (N = 8), and the remaining eight specimens obtained from the four premolars were allocated to the negative control group (N = 8). The specimens were then embedded in a round acrylic mold (outer diameter 20 mm, inner diameter 16 mm, height 8 mm) using self-cure acrylic resin (Ortho-jet Acrylic^c) with the cervical dentin exposed upward (Fig. 1).

After embedding the specimens, the exposed dentin surfaces were polished with abrasive papers (CC261^d) sized P100 to P1,200 to generate a reference surface meeting the ISO standard (ISO 6344). Polyvinylchloride (PVC) tape was attached onto each specimen to restrict and standardize the amount of exposed dentin (2 mm × 2 mm) to within the detection range of the surface profilometer (Surftest 402^e) (Fig. 1). Pre-treatment surface profiling was performed using the surface profilometer.

Table 1. Composition of the liquid dentifrice and conventional dentifrice.

Material/Brand	Dental abrasives	Composition
Liquid dentifrice/Garglin Gum-Guard	None	Cetylpyridinium chloride (USP) 50 mg, Tocopheryl acetate (USP) 200 mg, sodium monofluorophosphate (USP) 760 mg (equivalent to 1000 ppm of fluoride) per 100 g.
Conventional dentifrice/333 Clinic Total Care	Colloidal silicon dioxide 8 g (equivalent to 7.6 g of silicon dioxide), (equivalent to 8.7 g of silicon dioxide) per 100 g	Colloidal silicon dioxide (NF) 8 g, dental type silica (NF) 10 g, sodium monofluorophosphate (USP) 380 mg (equivalent to 500 ppm of fluoride), ocopheryl acetate (KP) T90 mg per 100 g.

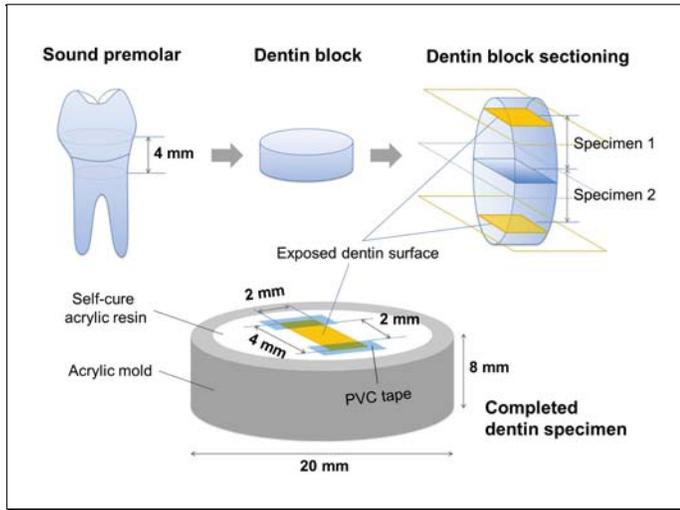


Fig. 1. Dentin specimen preparation and a representative completed specimen.

Table 2. Dentin wear after simulated toothbrushing with conventional dentifrice, liquid dentifrice, and distilled water.

Group	Conventional dentifrice	Liquid dentifrice	Distilled water
Ra (µm)	222.50 ± 74.52 ^a	12.47 ± 6.74 ^b	13.59 ± 6.61 ^b

^{a, b} Indicates a statistically significant difference between the groups (P < 0.05).

Dentin wear test - To prepare the conventional dentifrice, 25 g of 333 Clinic Total Care was mixed with 20 ml of distilled water on a magnetic stirrer according to the ISO standard. The liquid dentifrice (Garglin Gum-Guard) did not require additional preparation prior to the wear test. The pH values of the prepared conventional dentifrice and the liquid dentifrice were measured by a pH electrode (Orion Glass Body ROSS Combination pH Electrode⁵). A commercially available toothbrush (777 toothbrush⁶) was used in the simulated toothbrushing. The dentin specimens were submerged in the prepared dentifrices and distilled water (control group) to a depth of at least 3 mm. A total of 50,000 repeated strokes (2 strokes/second) were applied using the toothbrush onto each specimen under a 150 g-load.

After the surface wearing, the specimens were washed under running water, and post-treatment surface profiling was performed using the surface profilometer. Surface images of the specimen before and after wearing were superimposed and analyzed using the MTS^h 3D profiling system. The mean surface roughness (Ra) was calculated for each specimen (Table 2).

Scanning electron microscopy - A SEM was used to qualitatively assess the worn dentin surfaces. After coating the specimen with gold palladium in an ion sputtering coater, the worn dentin surfaces were imaged by SEM (S-4700 FESEM^l) at a 15.0-kV accelerating voltage and 12-mm working distance.

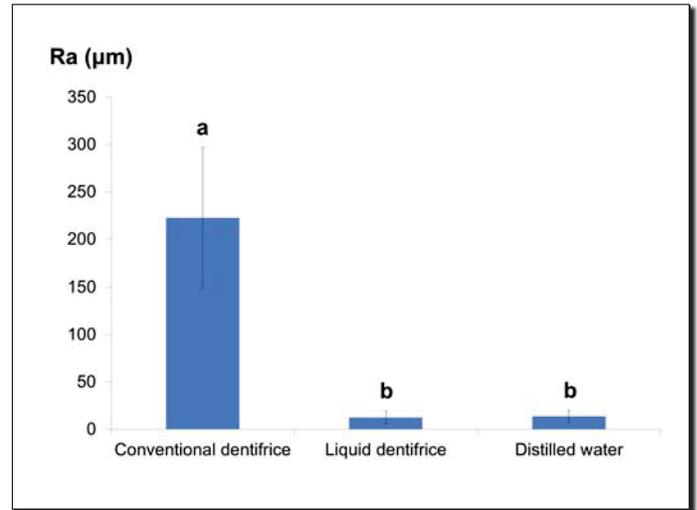


Fig. 2. Dentin wear after simulated toothbrushing with conventional dentifrice, liquid dentifrice, and distilled water. Statistically significant differences between the groups (P < 0.05) are indicated by superscripted letters.

Standard SEM images (×30 and ×300 magnifications) were obtained for each group.

Statistical analysis - The Ra values for the experimental and control groups were compared by one-way ANOVA, followed by Tukey's post-hoc test for multiple comparisons using IBM SPSS Statistics 20^l with an alpha of 5% (α = 0.05).

Results

pH measurement - The pH values were 6.93 ± 0.06 for the prepared conventional dentifrice and 6.70 ± 0.02 for the liquid dentifrice.

Mean surface roughness - The Ra values and the group comparisons are described in Table 2 and Fig. 2. The Ra values did not differ significantly between the liquid dentifrice and control groups (P > 0.05). However, the conventional dentifrice group showed a significantly higher Ra value compared to the Ra values in both the liquid dentifrice and control groups (P < 0.001). **SEM evaluation** - Figure 3 shows representative SEM images (×30 and ×300 magnifications) of the worn dentin specimens and reveals the differing surface topographies according to the dentifrice type. At ×30 magnification, the dentin specimens in the conventional dentifrice group exhibited crater-shaped defects along the worn surfaces with vertical dentin loss ranging from 50 to 200 µm (Fig. 3A). By contrast, the specimens in the liquid dentifrice and control groups did not exhibit any detectable vertical dentin loss along the worn surfaces (Figs. 3C, 3E). The conventional dentifrice group also showed characteristic furrow-like groove defects at a higher magnification (×300) parallel to the direc-

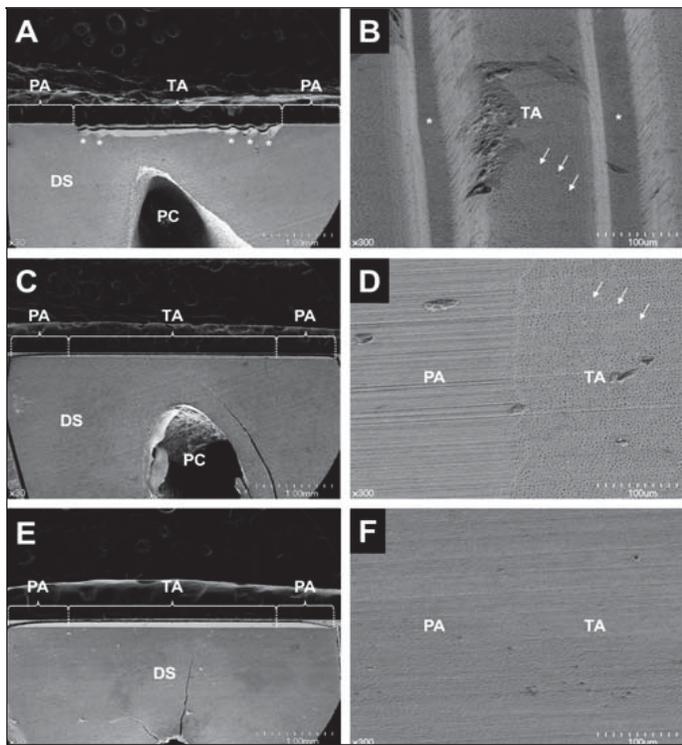


Fig. 3. Representative scanning electron microscope images of the worn dentin specimens treated with conventional dentifrice at **A**, $\times 30$ magnification and **B**, $\times 300$ magnification, liquid dentifrice at **C**, $\times 30$ magnification and **D**, $\times 300$ magnification, and distilled water at **E**, $\times 30$ magnification and **F**, $\times 300$ magnification. Micrographs in 3A, 3C, and 3E were taken in cross-sectional view, and those in 3B, 3D, and 3F were taken in routine two-dimensional view. Asterisks (3A and 3B) indicate the groove defects in the dentin specimen treated with conventional dentifrice. Arrows (3B and 3D) indicate open dentin tubule orifices. The left half of 3D and 3F includes the dentin surface protected using PVC tape during the wear process (PA). DS, dentin specimen; PC, pulp chamber; TA, toothbrushing area; PA, PVC-taped area.

tion of the simulated toothbrushing strokes (Fig. 3B). The liquid dentifrice and control groups showed flat and smooth worn surfaces (Figs. 3D, 3F). Dentin tubule exposure was apparent in both dentifrice groups (Figs. 3B, 3D) but was absent in the control group (Fig. 3F).

Discussion

This study evaluated and compared the effects of liquid dentifrice and conventional dentifrice on dentin wear and surface topography. When simulated toothbrushing (50,000 strokes) was performed *in vitro*, the liquid dentifrice caused significantly less dentin loss than the conventional dentifrice did, allowing us to reject the null hypothesis. This result is consistent with those of previous studies¹¹⁻¹⁵ reporting that dental abrasive is one of the main contributors to dentin wear caused by toothbrushing, because the conventional dentifrice in this study contained approximately 18 g of solid contents per 100 g of dentifrice, which are mostly dental abrasives, whereas the liquid dentifrice contained no dental abrasives and approximately 1 g of solid contents per 100 g of dentifrice. In this study, the dentin abrasivity of the dentifrices was compared with distilled water as the negative control, which has been widely used in previous studies,^{9,16,17} and notably the liquid dentifrice caused a similar degree of dentin wear compared to distilled water. Therefore, the liquid dentifrice in the present study appeared to be advantageous over a conventional denti-

frice in preventing NCCL caused by toothbrushing.

Cervical dentin loss not only reduces the thickness of the dentin surrounding the pulp, but also increases the number and diameter of exposed dentin tubules, which is closely associated with increased hydraulic conductance of dentin.¹⁸ Invasion of bacteria and bacterial by-products into the pulp may be facilitated, favoring development and progression of dentin hypersensitivity.⁴ With the SEM, the conventional dentifrice evaluated in this study caused considerable dentin loss and complete dentin tubule exposure along the worn dentin surfaces (Figs. 3A, 3B). Thus, dentifrices containing dental abrasives should be used with care to prevent dentin hypersensitivity, as well as NCCL. Regarding the dentin tubule exposure, liquid dentifrice revealed similar surface morphology compared to the conventional dentifrice. The dentin specimens treated with the liquid dentifrice showed open dentin tubules on the treated surface after toothbrushing, which is markedly different from the surface morphology of the specimens in the control group (Figs. 3D, 3F). Of the theories which explain the dentin tubule opening, the most popular one is the action of intraoral acid, which attacks and removes the smear layer especially when accompanied by mechanical stimulation.¹⁹ However, when considering that the dentifrices used in this study showed pH values which were almost neutral, this explanation appears to be less associated with this study. Meanwhile, previous studies^{20,21} suggested that the detergents contained in the dentifrice could be a factor which promotes the smear layer removal by reducing the abrasive resistance of the smear layer, which might be one of the causes of such alteration of surface morphology in the liquid dentifrice group. Although the liquid dentifrice was advantageous in minimizing the magnitude of dentin loss, its ability to remove the smear layer may increase the hydraulic conductance of dentin, tending to promote dentin sensitivity.

The plaque control ability of the dentifrice is another point to consider. Dental abrasives promote plaque removal during toothbrushing by increasing the abrasiveness of dentifrice; dentifrice with low abrasiveness was less efficient at plaque control.²² However, plaque control is affected by a variety of factors beyond the presence of dental abrasives such as the dentifrice composition and the brushing method. The liquid dentifrice in this study contained cetylpyridinium chloride (CPC), which reportedly functions as an antimicrobial agent,²³⁻²⁵ inhibits plaque formation,²⁶ and reduces previously formed plaque.²⁷ Another point is that both conventional and liquid dentifrice showed similar effectiveness in the smear layer removal when compared to the negative control (Fig. 3B, 3D, 3F), which implies their clinical potential in plaque control. Further studies evaluating the dentifrices regarding the ability to remove the biofilm would be helpful to compare the clinical benefits and limitations between the effects on the plaque control and the cervical dentin preservation.

In this study, the entire dentin wear process was performed in accordance with the ISO standards. However, while the ISO standard recommends 10,000 repeated strokes, the number of repeated strokes was increased to 50,000 because it was considered that 10,000 strokes, which is equivalent to 18.5 days of toothbrushing (assuming 1 stroke/second, 3 minutes/time, 3 times/day), was insufficient for reproducing daily toothbrushing. Therefore, in the present study, 50,000 strokes (equivalent to approximately 3 months of toothbrushing) were performed,

which is a more suitable simulation of daily toothbrushing performed over a long period.

The type of dentin substrate is another factor that significantly affects the results in dentin wear testing.²⁸ Sound teeth that were extracted due to orthodontic treatment were used in this study, and symmetrical dentin specimens from each tooth were randomly allocated to the liquid dentifrice and conventional dentifrice groups in order to minimize variation caused by the dentin substrates.

In this study, dentin wear from toothbrushing was reproduced in vitro, which provides a more controlled environment for quantitative analysis. However, many variables, such as the presence of saliva, oral biofilm and plaque deposition, hydrogen ion concentration (pH) and temperature changes, de- and remineralization of dentin, and individual brushing habits, influence the dentin wear process in vivo.^{11,29} Further laboratory studies and clinical trials are needed to address this particular study limitation.

In conclusion, within the limitations of this study, liquid dentifrice, which does not contain dental abrasives, caused less dentin wear than conventional dentifrice. However, both toothbrushing with liquid dentifrice and conventional dentifrice generated open dentin tubules on the dentin surface, which was not observed when toothbrushing was performed using distilled water. Additional laboratory studies and clinical trials are recommended to determine the clinical benefit of liquid dentifrice in mitigating dentin wear and dentin hypersensitivity.

- a. Dong-A Pharmaceutical, Seoul, Republic of Korea.
- b. JW Pharmaceutical, Seoul, Republic of Korea.
- c. Lang Dental Manufacturing Company, Wheeling, IL, USA.
- d. DEERFOS, Seoul, Republic of Korea.
- e. Mitutoyo, Kawasaki, Japan.
- f. Thermo Scientific, Beverly, MA, USA.
- g. LG Household & Health Care, Seoul, Republic of Korea.
- h. MTS Systems Corporation, Eden Prairie, MN, USA.
- i. Hitachi, Tokyo, Japan.
- j. SPSS Inc., Chicago, IL, USA.

Disclosure statement: The authors declared no conflict of interest. The first two authors contributed equally to this work and share first authorship. This work was supported by the Dong-A Pharmaceutical Research Fund and the Basic Science Research Program through the National Research Foundation of Korea (NRF), funded by the Ministry of Education, Science, and Technology (2012-009268).

Mr. Y. Jang is a graduate student, Dr. Roh is Professor, Department of Conservative Dentistry, College of Dentistry, Yonsei University, Seoul, Republic of Korea. Dr. Ihm is Research Professor, Office of Dental Education; Dr. Baik is a graduate student, Dental Research Institute, School of Dentistry, Seoul National University, Seoul, Republic of Korea. Ms. Yoo and Ms. D-H. Jang are graduate students, Department of Dental Hygiene, Wonju College of Medicine, Yonsei University, Wonju, Republic of Korea. Dr. Seo is Associate Professor, Department of Conservative Dentistry and Dental Research Institute, School of Dentistry, Seoul National University, Seoul, Republic of Korea.

References

1. Splieth CH, Tachou A. Epidemiology of dentin hypersensitivity. *Clin Oral Invest* 2013;17 Suppl 1:S3-S8.
2. Locker D, Grushka M. The impact of dental and facial pain. *J Dent Res* 1987;66:1414-1417.
3. Veitz-Keenan A, Barna JA, Strober B, Matthews AG, Collie D, Vena D, Curro FA, Thompson VP. Treatments for hypersensitive noncarious cervical lesions: A Practitioners Engaged in Applied Research and Learning (PEARL) Network randomized clinical effectiveness study. *J Am Dent Assoc* 2013;144:495-506.
4. West NX, Lussi A, Seong J, Hellwig E. Dentin hypersensitivity: Pain mechanisms and aetiology of exposed cervical dentin. *Clin Oral Invest* 2013;17 Suppl 1:S9-S19.
5. Aw TC, Lepe X, Johnson GH, Mancl L. Characteristics of noncarious cervical lesions: A clinical investigation. *J Am Dent Assoc* 2002;133:725-733.
6. Bergstrom J, Lavstedt S. An epidemiologic approach to toothbrushing and dental abrasion. *Community Dent Oral Epidemiol* 1979;7:57-64.
7. Addy M, Hunter ML. Can tooth brushing damage your health? Effects on oral and dental tissues. *Int Dent J* 2003;53 Suppl 3:177-186.
8. Dyer D, Addy M, Newcombe RG. Studies in vitro of abrasion by different manual toothbrush heads and a standard toothpaste. *J Clin Periodontol* 2000;27:99-103.
9. Turssi CP, Messias DC, Hara AT, Hughes N, Garcia-Godoy F. Brushing abrasion of dentin: Effect of diluent and dilution rate of toothpaste. *Am J Dent* 2010;23:247-250.
10. Addy M, Mostafa P, Newcombe RG. Dentine hypersensitivity: The distribution of recession, sensitivity and plaque. *J Dent* 1987;15:242-248.
11. Macdonald E, North A, Maggio B, Sufi F, Mason S, Moore C, Addy M, West NX. Clinical study investigating abrasive effects of three toothpastes and water in an in situ model. *J Dent* 2010;38:509-516.
12. Lewis R, Dwyer-Joyce RS, Pickles MJ. Interaction between toothbrushes and toothpaste abrasive particles in simulated tooth cleaning. *Wear* 2004;257:368-376.
13. Johannsen G, Tellefsen G, Johannsen A, Liljeborg A. The importance of measuring toothpaste abrasivity in both a quantitative and qualitative way. *Acta Odontol Scand* 2013;71:508-517.
14. Franzo D, Philpotts CJ, Cox TF, Joiner A. The effect of toothpaste concentration on enamel and dentine wear in vitro. *J Dent* 2010;38:974-979.
15. Giles A, Claydon NC, Addy M, Hughes N, Sufi F, West NX. Clinical in situ study investigating abrasive effects of two commercially available toothpastes. *J Oral Rehabil* 2009;36:498-507.
16. Lima JP, Melo MA, Passos VF, Braga CL, Rodrigues LK, Santiago SL. Dentin erosion by whitening mouthwash associated to toothbrushing abrasion: A focus variation 3D scanning microscopy study. *Microsc Res Tech* 2013;76:904-908.
17. De Menezes M, Turssi CP, Hara AT, Messias DC, Serra MC. Abrasion of eroded root dentine brushed with different toothpastes. *Clin Oral Invest* 2004;8:151-155.
18. Fogel HM, Marshall FJ, Pashley DH. Effects of distance from the pulp and thickness on the hydraulic conductance of human radicular dentin. *J Dent Res* 1988;67:1381-1385.
19. Eliades G, Mantzourani M, Labella R, Mutti B, Sharma D. Interactions of dentine desensitisers with human dentine: Morphology and composition. *J Dent* 2013;41 Suppl 4:S28-S39.
20. Addy M, Loynd T, Adams D. Dentine hypersensitivity - Effects of some proprietary mouthwashes on the dentine smear layer: A SEM study. *J Dent* 1991;19:148-152.
21. West N, Addy M, Hughes J. Dentine hypersensitivity: The effects of brushing desensitizing toothpastes, their solid and liquid phases, and detergents on dentine and acrylic: studies in vitro. *J Oral Rehabil* 1998;25:885-895.
22. Baxter PM, Davis WB, Jackson J. Toothpaste abrasive requirements to control naturally stained pellicle. The relation of cleaning power to toothpaste abrasivity. *J Oral Rehabil* 1981;8:19-26.
23. Furiga A, Dols-Lafargue M, Heyraud A, Chambat G, Lonvaud-Funel A, Badet C. Effect of antiplaque compounds and mouthrinses on the activity of glucosyltransferases from *Streptococcus sobrinus* and insoluble glucan production. *Oral Microbiol Immunol* 2008;23:391-400.
24. Hu D, Li X, Sreenivasan PK, DeVizio W. A randomized, double-blind clinical study to assess the antimicrobial effects of a cetylpyridinium chloride mouth rinse on dental plaque bacteria. *Clin Ther* 2009;31:2540-2548.
25. Steinberg D, Bachrach G, Gedalia I, Abu-Ata S, Rozen R. Effects of various antiplaque agents on fructosyltransferase activity in solution and immobilized onto hydroxyapatite. *Eur J Oral Sci* 2002;110:374-379.
26. Haps S, Slot DE, Berchier CE, Van der Weijden GA. The effect of cetylpyridinium chloride-containing mouth rinses as adjuncts to toothbrushing on plaque and parameters of gingival inflammation: A systematic review. *Int J Dent Hyg* 2008;6:290-303.
27. Silva MF, dos Santos NB, Stewart B, DeVizio W, Proskin HM. A clinical investigation of the efficacy of a commercial mouthrinse containing 0.05% cetylpyridinium chloride to control established dental plaque and gingivitis. *J Clin Dent* 2009;20:55-61.
28. Turssi CP, Messias DF, Corona SM, Serra MC. Viability of using enamel and dentin from bovine origin as a substitute for human counterparts in an intraoral erosion model. *Braz Dent J* 2010;21:332-336.
29. Han S, Lee E, Kim B, Jung H, Kwon H, Kim B. In situ study investigating abrasive effects of two different abrasive toothpastes. *J Korean Acad Oral Health* 2011;35:405-413.